



## Targets for Anti-HIV1- Agents as Personalized HIV Therapy: a Review Study

Mina Jafari<sup>1</sup>, Ava Shamipak<sup>1</sup>, Fateme Shahsavari<sup>1</sup>, Armin Nikdehqan<sup>1</sup>, Paniz Hajhossein<sup>1\*</sup>

<sup>1</sup>Department of Biology, Faculty of Basic Sciences, Islamic Azad University, East Tehran Branch, Tehran, Iran.

\*Corresponding author: Paniz Hajhossein, Department of Biology, Faculty of Basic Sciences, Islamic Azad University, East Tehran Branch, Tehran, Iran. Email: panizhajhossein@gmail.com.

DOI: [10.22034/pmj.2023.2011750.1016](https://doi.org/10.22034/pmj.2023.2011750.1016)

Submitted: 2023-08-23

Accepted: 2023-09-19

### Keywords:

HIV therapy  
Anti-HIV-1 Agents  
HIV medication resistance

©2023. Personalized Medicine Journal

### Abstract:

The enormous genetic variety of the viral population harbored by the patient and the large volume of therapeutic alternatives characterize HIV therapy. Each patient and period has its viral population. The enormous number of therapy possibilities makes selecting an ideal or near-optimal therapy challenging, especially among therapy-experienced patients. Over the last decade, computer-based medication selection that measures viral resistance to pharmaceuticals has become a norm for HIV patients. We explore the qualities of available systems and the field's viewpoints.

## INTRODUCTION

HIV is one of the most rapidly changing diseases known, and there is currently no HIV vaccine (1). Because the patient cannot be treated for the virus once infected, therapy aims to inhibit viral replication, alleviate symptoms, and extend life (2, 3). For this goal, more than two dozen distinct antiretroviral medications have been produced in record time for all other illnesses today (4, 5). Drugs inhibit a multitude of phases in the viral replication cycle (6-8). Although a particular medicine therapy can be beneficial for a long time, even years, the virus ultimately evolves into a resistant variety, resulting in therapeutic failure (9). When this occurs, a new treatment combination that effectively addresses the resistance profile displayed by the viral population currently in the patient must be chosen (10). This is a challenging undertaking, but appropriate tools can assist in selecting effective therapeutic alternatives for these individuals (11). This paper summarizes the history and current state of bioinformatics-based resistance analysis and future prospects (12).

### *HIV medication resistance assessment history*

There are two methods for HIV and other viral resistance analyses. Viruses are tested in vitro for sensitivity to various medicines in phenotypic resistance tests (13). This laboratory approach is highly informative in the context of research (14). However, it is unsuitable for clinical routine testing for

numerous reasons: the assay is challenging, requiring only a few highly specialized laboratories to do it, it is costly, and it takes a long time (more than a week) (15). Another option is genotypic resistance evaluation, which involves sequencing the relevant sections of the virus genome while analyzing the sequence concerning the virus's resistance phenotype (16). In industrialized nations, genotypic resistance screening is frequently used as a companion diagnosis in HIV therapy (17). The first attempt to analyse genotypic resistance information related to HIV in history was made utilising tables by expert committees that convened regularly (18). They made judgements based on evidence from the literature, laboratory data, and clinical (19). Regular updates to the resultant mutation lists were and continue to be released (20).

The mutation list has improved the efficacy of currently used antiretroviral treatments, but it has two shortcomings: The first is the table's minimal information content (21). A table, in particular, cannot convey relationships between alterations; instead, each mutation functions independently in giving the virus's resistance to the treatment, and neither the epistemic process nor desensitization is considered (22). The emergence of computerized rules-based systems has solved this constraint (23). In effect, they are sets of rules that can assume more sophisticated forms than the rules implied in the mutation tables (24). Consider the rule that says a virus is resistant to medication D if it possesses mutation M1 but not M2 (25). This

describes the virus's desensitization to medication D due to mutation M2 (26). The computer evaluates the relevant region of a viral genome versus all rules in the set in a rules-based system, also known as a resistance algorithm (27). There are numerous widely used systems, including those provided by the Stanford HIV Archive, Rega Institute, and ANRS (28). These techniques form the foundation of computer-based genotypic resistance data interpretation as additional testing for antiretroviral HIV medication selection (29).

#### *HIV-1 Life Cycle Factors as Anti-HIV-1 Agent Targets*

The HIV-1 life cycle is comprised of multiple phases, beginning with the adherence of an HIV-1 particle to the host cell membrane, where linkages between the HIV-1 envelope (gp120) and the cell surface CD4 receptor proceed by binding to the chemokine receptors CXCR4 or CCR5 (30). These contacts activate the HIV-1 fusion protein (gp41), producing cell-viral membrane fusion (31). The virion's contents are then released into the cytoplasm, where viral RNA is converted to double-stranded DNA by RNA-dependent DNA polymerase or HIV-1 reverse transcriptase (HIV-1-RT) (32).

Following that, viral DNA is incorporated into the host chromosome (33). Translation and transcription Using the cell's machinery, Gag and Gag-Pol polyproteins are converted into viral proteins and transported to the cell membrane, where virions are assembled, budded, and matured before being released as functional HIV-1 particles (34). In general, anti-HIV-1 medications should target viral or cellular proteins in the HIV-1 life cycle (35). Furthermore, interactions between such small compounds and target proteins should ideally result in HIV-1-specific inhibitory effects with minimal toxicity (36).

#### *Molecular docking of HIV protease Inhibitors*

Six authorized anti-HIV medications were chosen for testing (37). Although 3CLpro-2 and 3CLpro-1 have greater binding energies than all HIV protease inhibitor combinations used as positive controls, 3CLpro-2 has lower binding energy for all investigated inhibitors than its sibling 3CLpro-1 (38). This indicates that 3CLpro-2 has more remarkable binding affinities for inhibitors than 3CLpro-1 (39). Indinavir and darunavir have been shown to have a greater binding capacity to 3CLpro-2 than the other HIV protease inhibitors, and their interaction energy values are comparable to those of HIV inhibitors of protease (40).

When examined, the binding energy of the 3CLpro-2-darunavir complex (-10.24 kJ mol<sup>-1</sup>) is lower than that of its 3CLpro-2 indinavir equivalent (-10.02 kJ mol<sup>-1</sup>), showing that darunavir likely has a better affinity for 3CLpro-2 than indinavir (41). Because 3CLpro is required for coronavirus replication, the inhibitory action of these substances on 3CLpro-2 suggests

they might be used as anti-COVID-19 therapeutic medicines (42).

#### *New insights into the clinically validated antiretroviral targets*

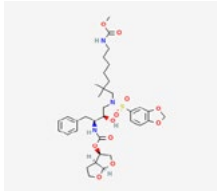
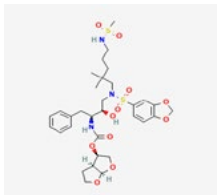
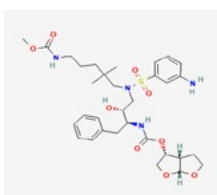
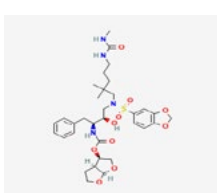
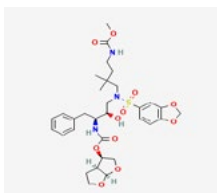
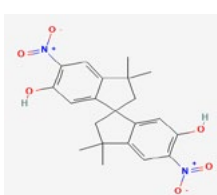
For the clinically validated HIV targets (RT, IN, PR, and CCR5), there is still significant scope for further development of novel inhibitors with distinct mechanisms of action, such as RNase H inhibitors, Nucleotide-competing RT inhibitors (NcRTIs), noncatalytic site (allosteric) IN inhibitors, and PR dimerization inhibitors. From the HIV therapy point of view (43), an allosteric inhibitor could restore the potency of an active site inhibitor against multidrug resistance. A combined therapy with an active site inhibitor and an allosteric inhibitor may be available as a new anti-HIV strategy to overcome drug resistance (44).

Recently, a high-resolution crystal structure of human CCR5 bound to the approved drug revealed a ligand-binding pocket that is distinct from the putative major binding sites for chemokines and HIV gp120, affording unprecedented insight into the mechanism of allosteric modulation of chemokine signalling and viral entry (45). This structure may suggest potential new drugs that could further inhibit the bioactivity of CCR5 (46). In addition, a subpocket on the N-trimer of HIV-1 gp41 was identified, with implications for developing anti-HIV entry inhibitors (47). Besides targeting an unconventional binding site, another rational design strategy to combat drug resistance has been to maximize highly conserved site interactions and significantly enhance extensive H-bond interactions with main-chain atoms strategy has been extensively employed to seek a variety (48).

#### *HIV Treatment*

Current anti-HIV medicines inhibit critical phases in the HIV life cycle; nevertheless, HIV can mutate, leaving these medications ineffective (49). HIV therapy is typically administered with two or three groups of ARVs, a process known as cART (50). ARVs are classified into five types: non-nucleoside reverse transcriptase drugs, protease inhibitors, entry/fusion inhibitors, integrase inhibitors, and nucleoside/nucleotide reverse transcriptase agents (51). The three medications of choice are an integrase-strand transfer blocker and two nucleoside reverse transcriptase inhibitors (52). ARVs are administered regularly, making adherence challenging (53). Any disruption in this everyday routine may result in the virus resurfacing (54). ARVs are administered orally; hence absorption is the primary method (55). Long-acting injectables (LAIs) such as Cabenuva, on the other hand, are injected intramuscularly rather than orally, giving LAIs an advantage over orally administered

**Table 1.** Substances retrieved from PubChem have been shown to inhibit HIV-1 protease in vitro.

Compound	Compound ID	Molecular Formula	Structure	HIV Protease activity
arylsulfonamide 15	CID480447	<a href="#">C<sub>35</sub>H<sub>49</sub>N<sub>3</sub>O<sub>11</sub>S</a>		Active
arylsulfonamide 16c	CID514961	<a href="#">C<sub>32</sub>H<sub>45</sub>N<sub>3</sub>O<sub>11</sub>S<sub>2</sub></a>		Active
arylsulfonamide 11b	CID480469	<a href="#">C<sub>32</sub>H<sub>46</sub>N<sub>3</sub>O<sub>9</sub>S</a>		Active
arylsulfonamide 16b	CID480440	<a href="#">C<sub>33</sub>H<sub>46</sub>N<sub>3</sub>O<sub>10</sub>S</a>		Active
arylsulfonamide 13	CID480441	<a href="#">C<sub>32</sub>H<sub>43</sub>N<sub>3</sub>O<sub>11</sub>S</a>		Active
CHEMBL60433	CID478338	<a href="#">C<sub>21</sub>H<sub>22</sub>N<sub>2</sub>O<sub>6</sub></a>		Inactive

medicines (56). The ARVs' biodistribution was also studied (57). New research, the first to evaluate ARV concentration from human brain tissues, found a greater concentration than any previously reported concentration (58). Furthermore, various ARVs might be more concentrated in different tissues, implying that particular phases in the HIV life cycle are not inhibited in specific reservoirs (59). As a result, ARV treatment considers medication-to-drug interactions, which may increase drug toxicity (60). Furthermore, some HIV patients use marijuana medically or recreationally, which can block the cytochrome P450 enzymes (61). This can eventually lead to increased ARV concentration in the circulation, which increases adverse effects and excretion rates (62).

#### Therapy prediction engines

A virtual phenotype is an estimate of the result of a laboratory experiment that serves as the foundation for selecting a suitable therapy in a second manual phase (63). The goal of therapy prediction engines is to automate the second stage (64). They rate various therapeutic alternatives in terms of their chance of success for a particular patient (65). Therapy prediction engines tackle a considerably more complex problem than virtual phenotypes since they try to predict clinical outcomes rather than merely a laboratory readout (66). The caretaker then chooses an appropriate therapy from the top-ranking treatments supplied by the prediction engine (67). In doing so, she will consider patient criteria the prediction engine does not evaluate, such as adverse reactions and ease of use (68).

The early treatment prediction engines constructed resistance ratings from virtual phenotypes relatively simply (69). Examples include the genotypic susceptibility score (GSS), a normalized sum of the virus's resistance ratings against several treatment types (70). More advanced systems use cutting-edge statistical learning methods to provide a prediction which involves both the estimated viral resistance and more details, such as drug interactions and an estimate

of the virus's expected evolutionary development to escape therapy in the future (71). Therapy forecasting systems can use the predictions provided by virtual phenotypes to predict therapy efficacy (72). Still, they can also use clinical correlates, information on patient history - such as previous use of drugs or combinations of drugs and previously observed resistance mutations - and even information on patient genotypes - such as HLA alleles (73). Several therapeutic prediction systems (THEO from the geno2pheno suite, the EuResist prognosis engine, and the RDI TREPS system) have been published and are available on the Internet (74). Furthermore, positions are under pressure from HLA presentation and certain antiviral medications (75).

#### CONCLUSIONS

Computer-assisted HIV treatment is at the forefront of personalized medicine. It is distinguished by complicated genomic biomarkers - essential portions of the viral genome - and a wide range of therapeutic alternatives. The therapy decision problem is dictated by viral resistance and is difficult, if possible, to solve manually. There are two versions of treatment selection systems. The first generation of virtual phenotypes predicts the virus's resistance to any given medicine in the arsenal. Virtual phenotypes are now used in clinical settings. The second generation of therapy prediction engines combines information about a patient, such as resistance estimations, patient history, and clinical correlations. Therapeutic prediction engines, which forecast the likelihood of therapeutic success, are the subject of much research. They are currently employed in research settings but have yet to reach clinical use.

The technique that has proven effective for HIV therapy can potentially be used to treat other infectious illnesses. A fast-increasing arsenal of antiviral medications is developing for HCV infection, leading to hepatitis C and hepatocellular cancer, and combination treatment therapy will become commonplace in the coming years. The geno2pheno

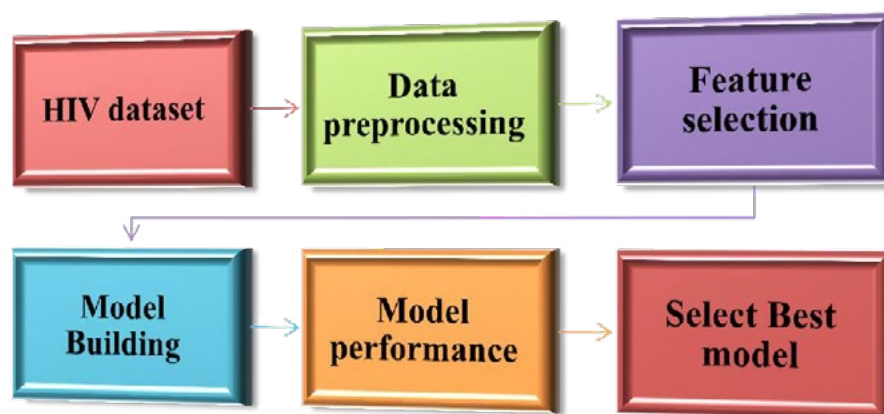


Fig 1. Flowchart of machine learning for HIV-failure prediction based on personalized medicine.

service already provides a virtual phenotype of HCV treatment resistance based on guidelines. With more phenotypic resistance data, we are prepared to give a mathematical model of drug resistance on that server. However, it is unknown if such a change is as essential for HCV as it is for HIV. There is optimism that individuals can quickly be cleansed of the virus using extremely efficient combination therapy treatments against HCV. This might reduce the requirement for computer-assisted therapy selection, as shown in TB when essential tabular criteria for medication administration suffice. Another situation in which this technique may be helpful is the HBV infection leading to hepatitis B, although its importance is unknown.

Using this kind of technology to combat tumors in the future has enormous promise. Cancer is similar to an HIV infection in that a parasite genome gains over the management of the cell, develops quickly, and escapes to resistant versions when challenged with medication therapy. The parasitic genome in cancer is that of the tumor cell. Compared to HIV, the genome and the pathways for resistance development are far more complicated and varied. Both situations share the problem caused by the variability of the parasite genome population. In this regard, the links between HIV and cancer are further examined.

#### Acknowledgements

The authors would like to thank the Biotechnology Research Centre staff members of the Parsian BioProducts Company (PBP) in Iran. This research received no specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

#### Conflict of Interest

The authors declare no conflict of interest.

#### Data Availability Statement

The data generated or analyzed during this study are included in this article.

#### REFERENCES

- Shah R, Mehta R. Why Covid-19 Vaccine Still not Invented to Relieve the Globe from Pandemic. *Journal of Applied Pharmacy*. 2021.
- Piri Gharaghie T, Hajimohammadi S. Comparison of anti-candida effects of aqueous, ethanolic extracts and essential oil of *E. angustifolia* with fluconazole on the growth of clinical strains of *Candida*. *New Cellular and Molecular Biotechnology Journal*. 2021 Jul 10;11(43):25-38.
- Piri Gharaghie T, Doosti A, Mirzaei SA. Detection of T6SS secretory system and membrane purine involved in antibiotic resistance in multidrug-resistant *Acinetobacter baumannii* isolates. *Journal of Microbial World*. 2021 May 22;14(1):47-58.
- Piri-Gharaghie T, Doosti A, Mirzaei SA. Novel adjuvant nano-vaccine induced immune response against *Acinetobacter baumannii*. *AMB Express*. 2023 Dec;13(1):1-6.
- Piri-Gharaghie T, Ghajari G, Hassanpoor M, Jegargoshe-Shirin N, Soosanirad M, Khayati S, Farhadi-Biregani A, Mirzaei A. Investigation of antibacterial and anticancer effects of novel niosomal formulated Persian Gulf Sea cucumber extracts. *Heliyon*. 2023 Mar 1;9(3).
- Poduri R, Joshi G, Jagadeesh G. Drugs targeting various stages of the SARS-CoV-2 life cycle: Exploring promising drugs for the treatment of Covid-19. *Cellular signalling*. 2020 Oct 1;74:109721.
- Taghiloo S, Ghajari G, Zand Z, Kabiri-Samani S, Kabiri H, Rajaei N, Piri-Gharaghie T. Designing Alginate/Chitosan Nanoparticles Containing *Echinacea angustifolia*: A Novel Candidate for Combating Multidrug-Resistant *Staphylococcus aureus*. *Chemistry & Biodiversity*. 2023 Jul;20(7):e202201008.
- Asadipour E, Asgari M, Mousavi P, Piri-Gharaghie T, Ghajari G, Mirzaei A. Nano-Biotechnology and Challenges of Drug Delivery System in Cancer Treatment Pathway. *Chemistry & Biodiversity*. 2023 Mar 1:e202201072.
- Coffin JM. HIV population dynamics in vivo: implications for genetic variation, pathogenesis, and therapy. *Science*. 1995 Jan 27;267(5197):483-9.
- Kieffer TL, Kwong AD, Picchio GR. Viral resistance to specifically targeted antiviral therapies for hepatitis C (STAT-Cs). *Journal of antimicrobial chemotherapy*. 2010 Feb 1;65(2):202-12.
- Schnipper LE, Davidson NE, Wollins DS, Tyne C, Blayney DW, Blum D, Dicker AP, Ganz PA, Hoverman JR, Langdon R, Lyman GH. American Society of Clinical Oncology statement: a conceptual framework to assess the value of cancer treatment options. *Journal of Clinical Oncology*. 2015 Aug 8;33(23):2563.
- Aranzana MJ, Decroocq V, Dirlwanger E, Eduardo I, Gao ZS, Gasic K, Iezzoni A, Jung S, Peace C, Prieto H, Tao R. *Prunus* genetics and applications after de novo genome sequencing: achievements and prospects. *Horticulture research*. 2019 Dec 1;6.
- Hirsch MS, Brun-Vézinet F, Clotet B, Conway B, Kuritzkes DR, D'Aquila RT, Demeter LM, Hammer SM, Johnson VA, Loveday C, Mellors JW. Antiretroviral drug resistance testing in adults infected with human immunodeficiency virus type 1: 2003 recommendations of an International AIDS Society-USA Panel. *Clinical Infectious Diseases*. 2003 Jul 1;37(1):113-28.
- Shanteau J. How much information does an expert use? Is it relevant?. *Acta psychologica*. 1992 Oct 1;81(1):75-86.
- McNerney R, Maeurer M, Abubakar I, Marais B, Mchugh TD, Ford N, Weyer K, Lawn S, Grobusch MP, Memish Z, Squire SB. Tuberculosis diagnostics and biomarkers: needs, challenges, recent advances, and opportunities. *Journal of Infectious Diseases*. 2012 May 15;205(suppl\_2):S147-58.
- Hirsch MS, Conway B, Richard TD, Johnson VA, Brun-Vézinet F, Clotet B, Demeter LM, Hammer SM, Jacobsen DM, Kuritzkes DR, Loveday C. Antiretroviral drug resistance testing in adults with HIV infection: implications for clinical management. *Jama*. 1998 Jun 24;279(24):1984-91.
- Bock C, Lengauer T. Managing drug resistance in cancer: lessons from HIV therapy. *Nature Reviews Cancer*. 2012 Jul;12(7):494-501.
- Kleinman, S., Busch, M.P., Murphy, E.L., Shan, H., Ness, P., Glynn, S.A. and National Heart, Lung, and Blood Institute. Personalized HIV therapy to control drug resistance. *Drug Discovery Today: Technologies*. 2014 Mar 1;11:57-64. *stitute Recipient Epidemiology and Donor*



- Evaluation Study (REDS-III), 2014. The National Heart, Lung, and Blood Institute Recipient Epidemiology and Donor Evaluation Study (REDS-III): a research program striving to improve blood donor and transfusion recipient outcomes. *Transfusion*, 54(3pt2), pp.942-955.
- 19.Lasater K. Clinical judgment development: Using simulation to create an assessment rubric. *Journal of nursing education*. 2007 Nov 1;46(11):496-503.
- 20.Bennett DE, Camacho RJ, Otelea D, Kuritzkes DR, Fleury H, Kiuchi M, Heneine W, Kantor R, Jordan MR, Schapiro JM, Vandamme AM. Drug resistance mutations for surveillance of transmitted HIV-1 drug-resistance: 2009 update. *PloS one*. 2009 Mar 6;4(3):e4724.
- 21.Aghemo A, De Francesco R. New horizons in hepatitis C antiviral therapy with direct-acting antivirals. *Hepatology*. 2013 Jul;58(1):428-38.
- 22.Creager AN, Gaudillière JP. Meanings in Search of Experiments and Vice-versa: the Invention of Allosteric regulation in Paris and Berkeley, 1959-1968. *Historical Studies in the Physical and Biological Sciences*. 1996 Jan 1;27(1):1-89.
- 23.Schätz B, Hölzl F, Lundkvist T. Design-space exploration through constraint-based model-transformation. In 2010 17<sup>th</sup> IEEE International conference and workshops on engineering of computer based systems 2010 Mar 22 (pp. 173-182). IEEE.
- 24.Vriend NJ. An illustration of the essential difference between individual and social learning, and its consequences for computational analyses. *Journal of economic dynamics and control*. 2000 Jan 1;24(1):1-9.
- 25.Van den Hoecke S, Ballegeer M, Vrancken B, Deng L, Job ER, Roose K, Schepens B, Van Hoecke L, Lemey P, Saelens X. In Vivo Therapy with M2e-Specific IgG Selects for an Influenza A Virus Mutant with Delayed Matrix Protein 2 Expression. *Mbio*. 2021 Aug 31;12(4):10-128.
- 26.Chizhnikov IV, Geraghty FM, Ogden DC, Hayhurst A, Antoniou M, Hay AJ. Selective proton permeability and pH regulation of the influenza virus M2 channel expressed in mouse erythrocyte cells. *The Journal of physiology*. 1996 Jul 15;494(2):329-36.
- 27.Raposo LM, Arruda MB, de Moraes Brindeiro R, Nobre FF. SIRA-HIV: A User-friendly System to Evaluate HIV-1 Drug Resistance from Next-generation Sequencing Data. In *BIOINFORMATICS 2020* (pp. 93-100).
- 28.Liu TF, Shafer RW. Web resources for HIV type 1 genotypic-resistance test interpretation. *Clinical infectious diseases*. 2006 Jun 1;42(11):1608-18.
- 29.Weinstein MC, Goldie SJ, Losina E, Cohen CJ, Baxter JD, Zhang H, Kimmel AD, Freedberg KA. Use of genotypic resistance testing to guide HIV therapy: clinical impact and cost-effectiveness. *Annals of internal medicine*. 2001 Mar 20;134(6):440-50.
- 30.Fanales-Belasio E, Raimondo M, Suligo B, Buttò S. HIV virology and pathogenetic mechanisms of infection: a brief overview. *Annali dell'Istituto superiore di sanita*. 2010;46:5-14.
- 31.Ji H, Shu W, Burling FT, Jiang S, Lu M. Inhibition of human immunodeficiency virus type 1 infectivity by the gp41 core: role of a conserved hydrophobic cavity in membrane fusion. *Journal of virology*. 1999 Oct 1;73(10):8578-86.
- 32.Hu WS, Hughes SH. HIV-1 reverse transcription. *Cold Spring Harbor perspectives in medicine*. 2012 Oct 1;2(10):a006882.
- 33.Schleper C, Kubo K, Zillig W. The particle SSV1 from the extremely thermophilic archaeon *Sulfolobus* is a virus: demonstration of infectivity and of transfection with viral DNA. *Proceedings of the National Academy of Sciences*. 1992 Aug 15;89(16):7645-9.
- 34.Sundquist WI, Kräusslich HG. HIV-1 assembly, budding, and maturation. *Cold Spring Harbor perspectives in medicine*. 2012 Mar 13:a006924.
- 35.Teixeira C, Gomes JR, Gomes P, Maurel F. Viral surface glycoproteins, gp120 and gp41, as potential drug targets against HIV-1: brief overview one quarter of a century past the approval of zidovudine, the first anti-retroviral drug. *European journal of medicinal chemistry*. 2011 Apr 1;46(4):979-92.
- 36.Mahajan SD, Aalinkel R, Law WC, Reynolds JL, Nair BB, Sykes DE, Yong KT, Roy I, Prasad PN, Schwartz SA. Anti-HIV-1 nanotherapeutics: promises and challenges for the future. *International journal of nanomedicine*. 2012 Oct 5:5301-14.
- 37.Sang P, Tian SH, Meng ZH, Yang LQ. Anti-HIV drug repurposing against SARS-CoV-2. *RSC advances*. 2020;10(27):15775-83.
- 38.Barghash RF, Fawzy IM, Chandrasekar V, Singh AV, Katha U, Mandour AA. In silico modeling as a perspective in developing potential vaccine candidates and therapeutics for COVID-19. *Coatings*. 2021 Oct 20;11(11):1273.
- 39.Schomburg D, Schomburg I. SARS coronavirus main proteinase 3.4. 22.69. Class 3.4-6 Hydrolases, Lyases, Isomerases, Ligases: EC 3.4-6. 2013:65-97.
- 40.Shivanika C, Kumar D, Rangunathan V, Tiwari P, Sumitha A. Molecular docking, validation, dynamics simulations, and pharmacokinetic prediction of natural compounds against the SARS-CoV-2 main-protease. *Journal of biomolecular structure & dynamics*. 2020:1.
- 41.Ibrahim MA, Abdelrahman AH, Allemailem KS, Almatroudi A, Moustafa MF, Hegazy ME. In silico evaluation of prospective anti-COVID-19 drug candidates as potential SARS-CoV-2 main protease inhibitors. *The Protein Journal*. 2021 Jun;40:296-309.
- 42.Nepali K, Sharma R, Sharma S, Thakur A, Liou JP. Beyond the vaccines: a glance at the small molecule and peptide-based anti-COVID19 arsenal. *Journal of Biomedical Science*. 2022 Sep 6;29(1):65.
- 43.Kang D, Chen W, Zhan P, Liu X. "Old Dogs with New Tricks": exploiting alternative mechanisms of action and new drug design strategies for clinically validated HIV targets. *Molecular BioSystems*. 2014;10(8):1998-2022.
- 44.Heredia A, Le N, Gartenhaus RB, Sausville E, Medina-Moreno S, Zapata JC, Davis C, Gallo RC, Redfield RR. Targeting of mTOR catalytic site inhibits multiple steps of the HIV-1 lifecycle and suppresses HIV-1 viremia in humanized mice. *Proceedings of the National Academy of Sciences*. 2015 Jul 28;112(30):9412-7.
- 45.De Amici M, Dallanocce C, Holzgrabe U, Tränkle C, Mohr K. Allosteric ligands for G protein-coupled receptors: A novel strategy with attractive therapeutic opportunities. *Medicinal research reviews*. 2010 May;30(3):463-549.
- 46.Blanpain C, Migeotte I, Lee B, Vakili J, Doranz BJ, Govaerts C, Vassart G, Doms RW, Parmentier M. CCR5 binds multiple CC-chemokines: MCP-3 acts as a natural antagonist. *Blood, The Journal of the American Society of Hematology*. 1999 Sep 15;94(6):1899-905.
- 47.Qiu Z, Chong H, Yao X, Su Y, Cui S, He Y. Identification

- and characterization of a subpocket on the N-trimer of HIV-1 Gp41: implication for viral entry and drug target. *Aids*. 2015 Jun 1;29(9):1015-24.
- 48.Tan JJ, Cong XJ, Hu LM, Wang CX, Jia L, Liang XJ. Therapeutic strategies underpinning the development of novel techniques for the treatment of HIV infection. *Drug discovery today*. 2010 Mar 1;15(5-6):186-97.
- 49.Rao KS, Ghorpade A, Labhasetwar V. Targeting anti-HIV drugs to the CNS. *Expert opinion on drug delivery*. 2009 Aug 1;6(8):771-84.
- 50.Heaton RK, Clifford DB, Franklin DR, Woods SP, Ake C, Vaida F, Ellis RJ, Letendre SL, Marcotte TD, Atkinson JH, Rivera-Mindt M. HIV-associated neurocognitive disorders persist in the era of potent antiretroviral therapy: CHARTER Study. *Neurology*. 2010 Dec 7;75(23):2087-96.
- 51.Clercq ED. New anti-HIV agents and targets. *Medicinal Research Reviews*. 2002 Nov;22(6):531-65.
- 52.Demarest J, Underwood M, St. Clair M, Dorey D, Brown D, Zolopa A. Dolutegravir-Based Regimens Are Active in Integrase Strand Transfer Inhibitor-Naïve Patients with Nucleoside Reverse Transcriptase Inhibitor Resistance. *AIDS Research and Human Retroviruses*. 2018 Apr 1;34(4):343-6.
- 53.Maskew M, MacPhail P, Menezes C, Rubel D. Lost to follow up—contributing factors and challenges in South African patients on antiretroviral therapy. *South African medical journal*. 2007 Oct 31;97(9):853-7.
- 54.Johansson O. Disturbance of the immune system by electromagnetic fields—A potentially underlying cause for cellular damage and tissue repair reduction which could lead to disease and impairment. *Pathophysiology*. 2009 Aug 1;16(2-3):157-77.
- 55.Sosnik A, Augustine R. Challenges in oral drug delivery of antiretrovirals and the innovative strategies to overcome them. *Advanced drug delivery reviews*. 2016 Aug 1;103:105-20.
- 56.Bassand C, Villosio A, Gianola L, Laue G, Ramazani F, Riebesehl B, Sanchez-Felix M, Sedo K, Ullrich T, Duvnjak Romc M. Smart design of patient-centric long-acting products: from preclinical to marketed pipeline trends and opportunities. *Expert opinion on drug delivery*. 2022 Oct 3;19(10):1265-83.
- 57.Kevadiya BD, Ottemann B, Mukadam IZ, Castellanos L, Sikora K, Hilaire JR, Machhi J, Herskovitz J, Soni D, Hasan M, Zhang W. Rod-shape theranostic nanoparticles facilitate antiretroviral drug biodistribution and activity in human immunodeficiency virus susceptible cells and tissues. *Theranostics*. 2020;10(2):630.
- 58.Asahchop EL, Meziane O, Mamik MK, Chan WF, Branton WG, Resch L, Gill MJ, Haddad E, Guimond JV, Wainberg MA, Baker GB. Reduced antiretroviral drug efficacy and concentration in HIV-infected microglia contributes to viral persistence in brain. *Retrovirology*. 2017 Dec;14:1-7.
- 59.Alam C, Whyte-Allman SK, Omeragic A, Bendayan R. Role and modulation of drug transporters in HIV-1 therapy. *Advanced drug delivery reviews*. 2016 Aug 1;103:121-43.
- 60.Tjagvad C. Addictive medication in relation to drug treatment and overdose death.
- 61.Andre M, Nair M, Raymond AD. HIV Latency and Nanomedicine Strategies for Anti-HIV Treatment and Eradication. *Biomedicines*. 2023 Feb 18;11(2):617.
- 62.Appay V, Sauce D. Immune activation and inflammation in HIV-1 infection: causes and consequences. *The Journal of Pathology: A Journal of the Pathological Society of Great Britain and Ireland*. 2008 Jan;214(2):231-41.
- 63.Wake DT, Smith DM, Kazi S, Dunnenberger HM. Pharmacogenomic clinical decision support: a review, how-to guide, and future vision. *Clinical Pharmacology & Therapeutics*. 2022 Jul;112(1):44-57.
- 64.Musen MA, Tu SW, Das AK, Shahar Y. EON: A component-based approach to automation of protocol-directed therapy. *Journal of the American Medical Informatics Association*. 1996 Nov 1;3(6):367-88.
- 65.Molitch ME, Beck JR, Dreisman M, Gottlieb JE, Paukerj SG. The cold thyroid nodule: an analysis of diagnostic and therapeutic options. *Endocrine reviews*. 1984 Apr 1;5(2):185-99.
- 66.Bender A, Cortés-Ciriano I. Artificial intelligence in drug discovery: what is realistic, what are illusions? Part 1: Ways to make an impact, and why we are not there yet. *Drug discovery today*. 2021 Feb 1;26(2):511-24.
- 67.Kazemi A, Salmani H, Shakibafard A, Fatehi F. New and emerging mobile technologies for healthcare (mHealth): A horizon scanning study. *Frontiers in Health Informatics*. 2019 Aug 21;8(1):17.
- 68.Kushniruk AW, Patel VL. Cognitive and usability engineering methods for the evaluation of clinical information systems. *Journal of biomedical informatics*. 2004 Feb 1;37(1):56-7
- 69.Belichenko PV, Kleschevnikov AM, Salehi A, Epstein CJ, Mobley WC. Synaptic and cognitive abnormalities in mouse models of Down syndrome: exploring genotype-phenotype relationships. *Journal of Comparative Neurology*. 2007 Oct 1;504(4):329-45.
- 70.Lengauer T, Pfeifer N, Kaiser R. Personalized HIV therapy to control drug resistance. *Drug Discovery Today: Technologies*. 2014 Mar 1;11:57-64
- 71.Stergiou KD, Minopoulos GM, Memos VA, Stergiou CL, Koidou MP, Psannis KE. A Machine Learning-Based Model for Epidemic Forecasting and Faster Drug Discovery. *Applied Sciences*. 2022 Oct 24;12(21):10766.
- 72.Sinha P, Kjelgaard MM, Gandhi TK, Tsourides K, Cardinaux AL, Pantazis D, Diamond SP, Held RM. Autism as a disorder of prediction. *Proceedings of the national academy of sciences*. 2014 Oct 21;111(42):15220-5.
- 73.Hirsch MS, Brun-Vézinet F, Clotet B, Conway B, Kuritzkes DR, D'Aquila RT, Demeter LM, Hammer SM, Johnson VA, Loveday C, Mellors JW. Antiretroviral drug resistance testing in adults infected with human immunodeficiency virus type 1: 2003 recommendations of an International AIDS Society-USA Panel. *Clinical Infectious Diseases*. 2003 Jul 1;37(1):113-28.
- 74.Vandamme AM, Camacho RJ, Ceccherini-Silberstein F, De Luca A, Palmisano L, Paraskevis D, Paredes R, Poljak M, Schmit JC, Soriano V, Walter H. European recommendations for the clinical use of HIV drug resistance testing: 2011 update. *Aids Rev*. 2011 Apr 1;13(2):77-108.
- 75.Gaudieri S, Rauch A, Pfafferoth K, Barnes E, Cheng W, McCaughan G, Shackel N, Jeffrey GP, Mollison L, Baker R, Furrer H. Hepatitis C virus drug resistance and immune-driven adaptations: relevance to new antiviral therapy. *Hepatology*. 2009 Apr;49(4):1069-82.