



# The Role of DNA Methylation in Development and Progression of Rheumatoid Arthritis

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DOI: 10.22034/pmj.2022.252438

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Submitted: 2021-12-10

Accepted: 2022-02-15

## Keywords:

Rheumatoid Arthritis  
DNA methylation  
Epigenetics  
Treatment

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## Abstract:

Rheumatoid arthritis (RA) is a chronic autoimmune disease of unknown etiology that results in progressive joint destruction and ultimately to disability. Currently effective biologic therapies, exist for approximately 40% of patients, but disease activity remains inadequately controlled in others. Therefore, it is crucial to identify specific markers that predict therapeutic response in various patients, prior to the initiation of therapy. DNA methylation, as an epigenetic factor, is increasingly being explored as a potential theranostic biomarker. It has been suggested that DNA methylation might contribute to RA development, nonetheless, with conflicting results. Epigenetic modules have provided a possible interface through which genetic and environmental risk factors contribute to the susceptibility and pathogenesis of RA. Hence, epigenetic regulators may provide promising drug targets to develop novel therapeutic drugs for tailored treatment of RA patients. Here we review the current knowledge regarding the role of DNA methylation in RA and indicate its potential therapeutic implications.

## INTRODUCTION

Rheumatoid arthritis (RA) is an autoimmune disease characterized by synovial hyperplasia and joint destruction (1). Its onset is progressive and invasive, which can lead to joint deformity and disability. The disease can cause a wide variety of symptoms, clinical forms and prognoses in patients (2). While its incidence begins to increase at the age of 25 years; it reaches a plateau at the age of 55. Furthermore, its prevalence is more than six times greater in women with 60 to 64 year-old than those in 18 to 29-year-old (3). In the industrialized world, RA affects more individuals than other countries. One third of people of all ages are also may affected by the disease at some point of their lifetime (4). The production of autoantibodies such as rheumatoid factor, and anticitrullinated protein antibodies (ACPA) are considered the hallmark of the disease that in turn supports an autoimmune etiology whereby an immune response is directed against an, as yet, unknown autoantigen. Although autoantibodies are an important characteristic of RA autoimmunity, some affected individuals may lack these autoantibodies in their blood (5, 6). The disease is complex and involves many environmental factors that trigger disease in

genetically susceptible individuals (7).

Major advancements in understanding the development of RA originate from studies investigating the expression and regulation of pro-inflammatory cytokines within the affected synovial tissue. Of these pro-inflammatory cytokines, tumor necrosis factor (TNF)- $\alpha$ , is a pivotal factor in the inflammatory cascade which led to its identification as a target for therapeutic intervention (8). TNF increases inflammation and tissue damage mediated by T cells, B cells, fibroblasts and macrophages in affected joints and also has systemic effects that can lead to comorbidities such as cardiovascular disease (9). Other cytokines such as IL-1 and IL-6 have similar roles in promoting the activation of T cells, B cells and osteoclasts while IL-17 promotes the infiltration of T cells and recruitment of monocytes and neutrophils, which also contribute to synovitis (10). B-lymphocytes are involved in the production of autoantibodies such as rheumatoid factor (RF) and antibodies against cyclic citrullinated peptide (anti-CCP). Differences in expression of anti-CCP and RF, rate of disease manifestation and variability of response to therapy cause heterogeneity of RA patients indicating, variation in pathophysiological

mechanisms, implication in the disease development and progression (11, 12).

In recent years, numerous studies have started to focus on the role of epigenetics in RA and investigate its contribution to the heterogeneity of patients (13). DNA methylation and histone modifications, are important epigenetic factors that affect the expression of immune-related genes and inflammation progression have become promising mechanisms to explain the pathogenesis of RA (14). Epigenetic changes in RA have been studied both in mononuclear cells of peripheral blood as well as other type of immune cells such as monocytes, T-cells and B-cells (15). In addition the epigenetic modifications in the RA synovial fibroblasts (RASFs) are of particular interest because of their aggressive phenotype, which remain stable for several passages in cell culture (16). RASFs are clue cells of joint damage and inflammation development due to pro-inflammatory and catabolic molecules synthesis, promoting abnormal proliferation and invasiveness (17). Numerous studies have found that methylation in immune cells may lead to RA progression through coordinated control of immune cell differentiation and function (18). In this review we aim to collate the current knowledge on DNA methylation in autoimmunity with a particular focus on RA, its role in altering gene expression in different cells that contribute to the pathogenesis of RA, and discuss its therapeutic and diagnostic potential.

#### *Molecular Mechanism of RA*

RA primarily affects synovial joints, in which the balance between recognition of pathogens and avoidance of self-attack is impaired and the immune system attacks and destroys healthy tissue (19). Additionally, there is increased recruitment and migration of immune cells from the bloodstream into the target tissue, including synovial membrane or synovial fluid (3, 20). Consequently, such an influx of activated immune cells producing an enhanced level of pro-inflammatory cytokines that leads to the progressive erosion of articular cartilage (21). Leukocytes, including T cells, B cells and phagocytes, are the main types of immune cells in the rheumatoid synovium (22). In fact, macrophages and granulocytes produce chemokines, pro-inflammatory cytokines, and reactive oxygen species, which are associated with classical inflammation. Besides, B lymphocytes play critical roles in the pathogenesis of RA (23). They are the main source of ACPAs and RF which involve in the formation of immune complexes as well as complement activation in the joints (24). B cells associated with the pathogenesis of RA disease not only activated by the presentation of antigens but also play a pivotal role in the development of the disease due to the production of antibodies, anti-self-antibodies

and cytokines. In addition to the role of soluble pro inflammatory molecules and activity markers, such as TNF, interleukin (IL)-6 and C-reactive protein, in the pathogenesis of RA, local synovial cellular interactions drive the key processes of long-term cellular proliferation and destruction of the rheumatoid joint (25, 26).

#### *influencing factors in RA*

People born with certain genes, called HLA class II genotypes, are more likely to develop RA (27). Although it is undeniable that genetic factors play the major role in the susceptibility of the illness, nonetheless, the low concordance rate (12–20%) observed in monozygotic twins suggests that environmental factors may also play a significant role in the pathogenesis of RA (28). Environmental factors such as smoking and infections may have affect the incidence of the disease as well as the rate of progression and severity of the RA (29). Moreover, other known environmental factors such as latent viral infections, sex hormones and deficiency of vitamin D may influence the disease (30). It is thought that these environmental factors influence epigenetic modifications, which in concert with the individual genetic susceptibility status result in the development of RA symptoms. Genetic heterogeneity however, does not explain all the features of illness (31, 32). Thus, investigation of epigenetic factors and mechanisms associated with the progression of the disease and response to treatment is increasingly important. Nevertheless, Investigating the epigenetic landscape can provide novel therapeutic targets (5, 33).

#### *Treatment for arthritis rheumatoid*

The main treatment goals are to control inflammation, ease pain, and reduce disability linked to RA (34). Current treatment guidelines recommend that patients initially treated with a combination of corticosteroids and disease-modifying antirheumatic drugs (DMARDs) to slow down disease progression and reduce synovitis along with disability (35). Though many people with RA need to take more than one drug to combat the disease. This is because drugs work in different ways to reduce the symptoms (36). There are three types of DMARD:

- conventional synthetic DMARDs (sometimes called csDMARDs)
- biological therapies (sometimes called bDMARDs).
- targeted synthetic DMARDs (sometimes called tsDMARDs)

Some DMARDs include hydroxychloroquine (Plaquenil), leflunomide (Arava), sulfasalazine (Azulfidine), or tofacitinib (Xeljanz) (37, 38). Although steroids are sometimes known by their full name: corticosteroids, it helps to reduce the pain, stiffness and inflammation caused by RA (39). The most common anti-inflammatory steroids

include hydrocortisone (Cortef), methylprednisolone (Medrol), and prednisone (Deltasone) (40). Non-steroidal anti-inflammatory drugs (NSAIDs) can be used to help control symptoms of pain, swelling or stiffness. They commonly used in combination with painkillers (41).

Biologic therapies are genetically engineered human proteins that specifically target inflammatory cytokines, such as TNF- $\alpha$  and IL-6 or immune pathways, such as CTLA-4 costimulatory pathways, or B cells (42). Though when bone damage from the arthritis become severe or pain is not controlled with medications, surgery is an option to restore function to a damaged joint (43).

#### Mechanism of DNA methylation

Epigenetics refers to chemical modifications that influence gene regulation without changing the DNA sequence (44). These alterations include DNA methylation and post-translational modifications of histone proteins(45). This review will focus mainly on studies of DNA methylation as a biomarker of response to treatment in RA. DNA methylation is a heritable epigenetic marker involving the covalent transfer of a methyl group to the C-5 position of the cytosine ring of DNA by DNA methyltransferases (Fig1) (4, 46). However, more than 98% of DNA methylation occurs in a CpG dinucleotide context in somatic cells, while as much as a quarter of all methylation appears in a non-CpG context in embryonic stem cells (ESCs) (47).

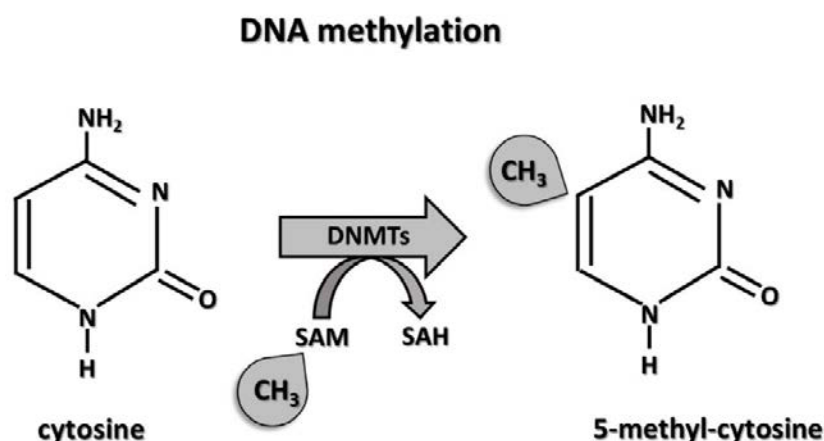
DNA methylation is a more stable biomarker than gene expression, and aberrant methylation has been reported in several cancers (48). DNA methylation has also been found to predict response to therapy; for example, an epigenome-wide association study (EWAS) identified

methylation signatures as a predictive response to anti-EGFR, a common therapeutic for metastatic colorectal cancers (49, 50).

#### Impaired DNA methylation in RA

Whole genome analysis of aged individuals has highlighted a number of hypomethylated regions that may contribute to age-related diseases, including some autoimmune diseases such as RA (51). The first evidence to suggest that DNA methylation may play a role in aging and autoimmunity came from studies investigating the effect of the DNA methyltransferase inhibitor 5-azacytidine that can induce symptoms associated with autoimmunity (52). There is emerging evidence of the interrelationship between DNA methylation and inflammation in regulating immune pathways. For example, the cytokine, IL-6, has been reported to increase the expression of DNMT1, levels of which correlate with DNA methylation in T cells (53). Studies have shown that the extent of methylation regulates migration, differentiation, and activation of T-cells. T cell activation leads to demethylation of the interleukin-2 promoter, resulting in interleukin-2 production (54). Such impairment of DNA methylation may play a role in RA pathogenesis. Specifically for RA, differential DNA methylation has been demonstrated in the IL-6 promoter (55). In 2008, Nile and colleagues found that IL-6 promoter methylation reduced transcriptional activity and identified a single CpG within the IL-6 promoter that was key to regulating IL-6 gene expression (53).

Recent studies confirmed a global DNA hypomethylation in T-cells and monocytes of RA patients compared to healthy individuals (56). In CD4+ T cells, 383 hyper- and 785 hypo-methylated genes were identified in RA patients ( $p < 3.4 \times 10^{-7}$ ), including three regions within HLA that were frequently



**Fig1.** DNA methyltransferases (DNMTs), responsible for the transfer of a methyl group from the universal methyl donor, S-adenosyl-L-methionine (SAM), to the 5-position of cytosine residues in DNA (46).

hypomethylated (57). Genome-wide analysis of DNA methylation by microarrays revealed its impact in B-cells on the early stages of patients who have not yet received treatment compared to healthy donors. Genome-wide methylation change was also found in T and B (58). A study identified 150 and 113 CpG loci with unique methylation characteristics in T and B lymphocytes in patients with ERA.

Rheumatoid arthritis fibroblast-like synoviocytes (RA-FLS) are involved in the release of inflammatory mediators and matrix degrading enzymes, which are key effector cells leading to synovial inflammation and destruction of bone and cartilage (59). The changes in DNA methylation in RA-FLS play important roles in the pathogenesis of illness. Hypomethylation in RA-FLS may be caused by the downregulation of DNMT1 and DNMT3A after inflammatory environmental stimulation (60).

#### *DNA methylation as a biomarker in RA*

Biologic drug therapies represent a huge advance in the treatment of RA (61). However, effective treatment of disease is achieved in only 30% of patients, making identification of biomarkers of response a research priority. Since DNA methylation appears to have a role in RA pathogenesis, it may also be a suitable biomarker of treatment response (62). Just as serologic and genetic studies indicate that there may be more than one sub-type of RA with a wide range of responses to biologic treatment, differences in baseline methylation status may also serve as a marker of varied disease subtypes that might respond better to therapies targeting the particular pathway involved (63). DNA methylation may thus provide a biomarker of subsequent treatment response (64). It is urgent to find novel markers to augment the diagnostic accuracy, prediction of disease onset, and its progression. For example Methylation levels of SHROOM1 in ERA are substantially increased, hence it can be applied as an early diagnostic biomarker (65). Additionally, identification of aberrant DNA methylation may change disease onset which in turn it might lead to a better understanding of the risk factors that contribute to disease development and thus result in the identification of specific biomarkers for disease analysis(66). Recently, reverse transcriptase (RT)-PCR assays have been developed to quantify the number of Foxp3+ cells within RA tissue samples (67).

It should be noted that there may be other appropriate biomarkers of response. Micro-RNAs (miRNA) are small, noncoding RNA structures that act as regulators of gene expression (68). There is increasing evidence that implicates dysregulation of miRNA in blood, T cells, and synovial fibroblasts in inflammation and joint destruction are found in RA patients. There have been a number of successful therapies for patients,

however, a proportion of patients fail to respond to conventional therapy (69). Ideally, it would be useful to identify this fraction of nonresponders earlier in the course of the disease, to provide better treatment regimes that are tailored towards the individual patient. It is becoming clear that RA patients display a differentially methylated genome when compared to healthy individuals (70). This raises the possibility that measuring DNA methylation patterns of responders and nonresponders may lead to the use of DNA methylation as a predictive biomarker for treatment response (71).

## CONCLUSIONS

In recent decades, many studies have shown that epigenetic mechanisms are involved in the regulation of all biological processes in the body from impregnation to death. In recent years, a major advancement has taken place in understanding the role of DNA methylation in the pathogenesis of RA, hence it can be used as effective biomarker in the disease process (72). It is hoped that the progress made in identifying epigenetic mechanisms occurring in cancer can also be exploited in inflammatory disease for other disease assessment. Unfortunately, studies on the clinical use of epigenetic drugs modulating aberrant DNA methylation patterns in RA are at a very early stage. More research should be conducted on DNA methylation in regard to treatment and diagnosis of cancer and proliferative diseases (73). Moreover, identified differential methylation genes can be applied as useful biomarkers to predict disease progression and severity and also provide potential therapeutic targets for RA. Epigenetic modifications as drug targets could provide a new direction of pharmacological research for the development of novel drugs that alleviate symptoms of high toxicity, low efficiency, and high cost of the current medical care. For example, demethylation of FOXP3 is used as a biomarker to evaluate the therapeutic drug response, which provides a direction for the precision treatment of RA (74). It is crucial to find new DNA methylation biomarkers that can be used in everyday practice to detect early onset of RA before the induction of irreversible joint destruction. These knowledges collectively may not only delay or reduce disease progression but also it decreases the costs of health care (75).

## REFERENCES

1. McInnes IB, Schett G. The pathogenesis of rheumatoid arthritis. *New England Journal of Medicine*. 2011 Dec 8;365(23):2205-19.
2. Burmester GR, Pope JE. Novel treatment strategies in rheumatoid arthritis. *The Lancet*. 2017 Jun 10;389(10086):2338-48.
3. Feldmann M, Maini RN. Anti-TNF alpha therapy of rheumatoid arthritis: what have we learned? *Annu. Rev. Immunol.* 19, 163–196 (2001).

4. Aletaha D, Smolen JS. Diagnosis and management of rheumatoid arthritis: a review. *Jama*. 2018 Oct 2;320(13):1360-72.
5. McInnes IB, Schett G. Pathogenetic insights from the treatment of rheumatoid arthritis. *The Lancet*. 2017 Jun 10;389(10086):2328-37.
6. Okada Y, Eyre S, Suzuki A, Kochi Y, Yamamoto K. Genetics of rheumatoid arthritis: 2018 status. *Annals of the rheumatic diseases*. 2019 Apr 1;78(4):446-53.
7. Ai R, Laragione T, Hammaker D, Boyle DL, Wildberg A, Maeshima K, Palescandolo E, Krishna V, Pocalyko D, Whitaker JW, Bai Y. Comprehensive epigenetic landscape of rheumatoid arthritis fibroblast-like synoviocytes. *Nature communications*. 2018 May 15;9(1):1-1.
8. Tseng CC, Lin YZ, Lin CH, Li RN, Tsai WC, Ou TT, Wu CC, Sung WY, Yen JH. Genetic and epigenetic alteration of the programmed cell death 1 in rheumatoid arthritis. *European Journal of Clinical Investigation*. 2019 Oct;49(10):e13094.
9. Meng H, Cao Y, Qin J, Song X, Zhang Q, Shi Y, Cao L. DNA methylation, its mediators and genome integrity. *International journal of biological sciences*. 2015;11(5):604.
10. Mazzone R, Zwergel C, Artico M, Taurone S, Ralli M, Greco A, Mai A. The emerging role of epigenetics in human autoimmune disorders. *Clinical epigenetics*. 2019 Dec;11(1):1-5.
11. Qiu H, Wu H, Chan V, Lau CS, Lu Q. Transcriptional and epigenetic regulation of follicular T-helper cells and their role in autoimmunity. *Autoimmunity*. 2017 Feb 17;50(2):71-81.
12. Bullock J, Rizvi SA, Saleh AM, Ahmed SS, Do DP, Ansari RA, Ahmed J. Rheumatoid arthritis: a brief overview of the treatment. *Medical Principles and Practice*. 2018;27(6):501-7.
13. Tam LS, Gu J, Yu D. Pathogenesis of ankylosing spondylitis. *Nature Reviews Rheumatology*. 2010 Jul;6(7):399-405.
14. Glant TT, Mikecz K, Rauch TA. Epigenetics in the pathogenesis of rheumatoid arthritis. *BMC medicine*. 2014 Dec;12(1):1-5.
15. Ospelt C. Epigenetic biomarkers in rheumatology—the future?. *Swiss Medical Weekly*. 2016;146:w14312.
16. Hardy RS, Hülso C, Liu Y, Gasparini SJ, Fong-Yee C, Tu J, Stoner S, Stewart PM, Raza K, Cooper MS, Seibel MJ. Characterisation of fibroblast-like synoviocytes from a murine model of joint inflammation. *Arthritis Research & Therapy*. 2013 Feb;15(1):1-5.
17. Lefevre S, Meier FM, Neumann E, Muller-Ladner U. Role of synovial fibroblasts in rheumatoid arthritis. *Current pharmaceutical design*. 2015 Jan 1;21(2):130-41.
18. Nasonov EL, Lila AM. Rheumatoid arthritis: achievements and unresolved issues. *Terapevticheskiy arkhiv*. 2019 May 15;91(5):4-7.
19. Hitchon CA, El-Gabalawy HS. Suppl 1: the synovium in rheumatoid arthritis. *The open rheumatology journal*. 2011;5:107.
20. Laria A, Lurati A, Marrazza M, Mazzocchi D, Re KA, Scarpellini M. The macrophages in rheumatic diseases. *Journal of inflammation research*. 2016;9:1.
21. Ciechomska M, Wilson CL, Floudas A, Hui W, Rowan AD, van Eden W, Robinson JH, Knight AM. Antigen-specific B lymphocytes acquire proteoglycan aggrecan from cartilage extracellular matrix resulting in antigen presentation and CD 4+ T-cell activation. *Immunology*. 2014 Jan;141(1):70-8.
22. Bogdanos DP, Smyk DS, Rigopoulou EI, Mytilinaiou MG, Heneghan MA, Selmi C, Gershwin ME. Twin studies in autoimmune disease: genetics, gender and environment. *Journal of autoimmunity*. 2012 May 1;38(2-3):J156-69.
23. Shi J, van Steenberg HW, van Nies JA, Levarht EN, Huizinga TW, van der Helm-van AH, Toes RE, Trouw LA. The specificity of anti-carbamylated protein antibodies for rheumatoid arthritis in a setting of early arthritis. *Arthritis research & therapy*. 2015 Dec;17(1):1-6.
24. Ciechomska M, O'Reilly S. Epigenetic modulation as a therapeutic prospect for treatment of autoimmune rheumatic diseases. *Mediators of inflammation*. 2016 Aug 10;2016.
25. Carmona-Rivera C, Carlucci PM, Moore E, Lingampalli N, Uchtenhagen H, James E, Liu Y, Bicker KL, Wahamaa H, Hoffmann V, Catrina AI. Synovial fibroblast-neutrophil interactions promote pathogenic adaptive immunity in rheumatoid arthritis. *Science immunology*. 2017 Apr 14;2(10):eaag3358.
26. Klein K, Kabala PA, Grabiec AM, Gay RE, Kolling C, Lin LL, Gay S, Tak PP, Prinjha RK, Ospelt C, Reedquist KA. The bromodomain protein inhibitor I-BET151 suppresses expression of inflammatory genes and matrix degrading enzymes in rheumatoid arthritis synovial fibroblasts. *Annals of the rheumatic diseases*. 2016 Feb 1;75(2):422-9.
27. Abbasi M, Mousavi MJ, Jamalzehi S, Alimohammadi R, Bezvan MH, Mohammadi H, Aslani S. Strategies toward rheumatoid arthritis therapy; the old and the new. *Journal of cellular physiology*. 2019 Jul;234(7):10018-31.
28. Frisell T, Saevarsdottir S, Askling J. Family history of rheumatoid arthritis: an old concept with new developments. *Nature Reviews Rheumatology*. 2016 Jun;12(6):335-43.
29. Conigliaro P, D'Antonio A, Pinto S, Chimenti MS, Triggianese P, Rotondi M, Perricone R. Autoimmune thyroid disorders and rheumatoid arthritis: A bidirectional interplay. *Autoimmunity Reviews*. 2020 Jun 1;19(6):102529.
30. van der Woude D, van der Helm-van AH. Update on the epidemiology, risk factors, and disease outcomes of rheumatoid arthritis. *Best Practice & Research Clinical Rheumatology*. 2018 Apr 1;32(2):174-87.
31. van der Woude D, van der Helm-van AH. Update on the epidemiology, risk factors, and disease outcomes of rheumatoid arthritis. *Best Practice & Research Clinical Rheumatology*. 2018 Apr 1;32(2):174-87.
32. Suzuki A, Kochi Y, Okada Y, Yamamoto K. Insight from genome-wide association studies in rheumatoid arthritis and multiple sclerosis. *FEBS letters*. 2011 Dec 1;585(23):3627-32.
33. Lundberg K, Wegner N, Yucel-Lindberg T, Venables PJ. Periodontitis in RA—the citrullinated enolase connection. *Nature Reviews Rheumatology*. 2010 Dec;6(12):727-30.
34. Singh JA, Saag KG, Bridges Jr SL, Akl EA, Bannuru RR, Sullivan MC, Vaysbrot E, McNaughton C, Osani M, Shmerling RH, Curtis JR. 2015 American College of Rheumatology guideline for the treatment of rheumatoid arthritis. *Arthritis & rheumatology*. 2016 Jan;68(1):1-26.
35. Scott DL. Biologics-based therapy for the treatment of rheumatoid arthritis. *Clinical Pharmacology & Therapeutics*. 2012 Jan;91(1):30-43.
36. Burmester GR, Pope JE. Novel treatment strategies in rheumatoid arthritis. *The Lancet*. 2017 Jun 10;389(10086):2338-48.
37. McInnes IB, Schett G. Pathogenetic insights from the treatment of

- rheumatoid arthritis. *The Lancet*. 2017 Jun 10;389(10086):2328-37.
38. Bullock J, Rizvi SA, Saleh AM, Ahmed SS, Do DP, Ansari RA, Ahmed J. Rheumatoid arthritis: a brief overview of the treatment. *Medical Principles and Practice*. 2018;27(6):501-7.
  39. Oliver J, Plant D, Webster AP, Barton A. Genetic and genomic markers of anti-TNF treatment response in rheumatoid arthritis. *Biomarkers in Medicine*. 2015 Jun;9(6):499-512.
  40. Rein P, Mueller RB. Treatment with biologicals in rheumatoid arthritis: an overview. *Rheumatology and therapy*. 2017 Dec;4(2):247-61.
  41. Zampeli E, Vlachoyiannopoulos PG, Tzioufas AG. Treatment of rheumatoid arthritis: unraveling the conundrum. *Journal of autoimmunity*. 2015 Dec 1;65:1-8.
  42. Taylor PC, Abdul Azeez M, Kiriakidis S. Filgotinib for the treatment of rheumatoid arthritis. *Expert Opinion on Investigational Drugs*. 2017 Oct 3;26(10):1181-7.
  43. Kerschbaumer A, Sepriano A, Smolen JS, van der Heijde D, Dougados M, van Vollenhoven R, McInnes IB, Bijlsma JW, Burmester GR, de Wit M, Falzon L. Efficacy of pharmacological treatment in rheumatoid arthritis: a systematic literature research informing the 2019 update of the EULAR recommendations for management of rheumatoid arthritis. *Annals of the rheumatic diseases*. 2020 Jun 1;79(6):744-59.
  44. Stricker SH, Köferle A, Beck S. From profiles to function in epigenomics. *Nature Reviews Genetics*. 2017 Jan;18(1):51-66.
  45. Marchal C, Miotto B. Emerging concept in DNA methylation: role of transcription factors in shaping DNA methylation patterns. *Journal of cellular physiology*. 2015 Apr;230(4):743-51.
  46. Nair N, Wilson AG, Barton A. DNA methylation as a marker of response in rheumatoid arthritis. *Pharmacogenomics*. 2017 Sep;18(14):1323-32.
  47. Renauer P, Coit P, Jeffries MA, Merrill JT, McCune WJ, Maksimowicz-McKinnon K, Sawalha AH. DNA methylation patterns in naïve CD4+ T cells identify epigenetic susceptibility loci for malar rash and discoid rash in systemic lupus erythematosus. *Lupus science & medicine*. 2015 Sep 1;2(1):e000101.
  48. Blanch M, Mosquera JL, Ansoleaga B, Ferrer I, Barrachina M. Altered mitochondrial DNA methylation pattern in Alzheimer disease-related pathology and in Parkinson disease. *The American journal of pathology*. 2016 Feb 1;186(2):385-97.
  49. Ouchi K, Takahashi S, Yamada Y, Tsuji S, Tatsuno K, Takahashi H, Takahashi N, Takahashi M, Shimodaira H, Aburatani H, Ishioka C. DNA methylation status as a biomarker of anti-epidermal growth factor receptor treatment for metastatic colorectal cancer. *Cancer science*. 2015 Dec;106(12):1722-9.
  50. Nervi C, De Marinis E, Codacci-Pisanelli G. Epigenetic treatment of solid tumours: a review of clinical trials. *Clinical epigenetics*. 2015 Dec;7(1):1-20.
  51. Cribbs A, Feldmann M, Oppermann U. Towards an understanding of the role of DNA methylation in rheumatoid arthritis: therapeutic and diagnostic implications. *Therapeutic advances in musculoskeletal disease*. 2015 Oct;7(5):206-19.
  52. Jin Z, Liu Y. DNA methylation in human diseases. *Genes & diseases*. 2018 Mar 1;5(1):1-8.
  53. Sun B, Hu L, Luo ZY, Chen XP, Zhou HH, Zhang W. DNA methylation perspectives in the pathogenesis of autoimmune diseases. *Clinical Immunology*. 2016 Mar 1;164:21-7.
  54. de Andres MC, Perez-Pampin E, Calaza M, Santaclara FJ, Ortea I, Gomez-Reino JJ, Gonzalez A. Assessment of global DNA methylation in peripheral blood cell subpopulations of early rheumatoid arthritis before and after methotrexate. *Arthritis research & therapy*. 2015 Dec;17(1):1-9.
  55. Ishida K, Kobayashi T, Ito S, Komatsu Y, Yokoyama T, Okada M, Abe A, Murasawa A, Yoshie H. Interleukin-6 gene promoter methylation in rheumatoid arthritis and chronic periodontitis. *Journal of periodontology*. 2012 Jul;83(7):917-25.
  56. Park SH, Kim SK, Choe JY, Moon Y, An S, Park MJ, Kim DS. Hypermethylation of EBF3 and IRX1 genes in synovial fibroblasts of patients with rheumatoid arthritis. *Molecules and cells*. 2013 Apr;35(4):298-304.
  57. Hammaker D, Whitaker JW, Maeshima K, Boyle DL, Ekwall AK, Wang W, Firestein GS. LBH gene transcription regulation by the interplay of an enhancer risk allele and DNA methylation in rheumatoid arthritis. *Arthritis & rheumatology*. 2016 Nov;68(11):2637-45.
  58. Guderud K, Sunde LH, Flåm ST, Mæhlen MT, Mjaavatten MD, Lillegraven S, Aga AB, Evenrød IM, Norli ES, Andreassen BK, Franzenburg S. Rheumatoid Arthritis Patients, Both Newly Diagnosed and Methotrexate Treated, Show More DNA Methylation Differences in CD4+ Memory Than in CD4+ Naïve T Cells. *Frontiers in immunology*. 2020:194.
  59. Guo S, Zhu Q, Jiang T, Wang R, Shen Y, Zhu X, Wang Y, Bai F, Ding Q, Zhou X, Chen G. Genome-wide DNA methylation patterns in CD4+ T cells from Chinese Han patients with rheumatoid arthritis. *Modern rheumatology*. 2017 May 4;27(3):441-7.
  60. Sellars M, Huh JR, Day K, Issuree PD, Galan C, Gobeil S, Absher D, Green MR, Littman DR. Regulation of DNA methylation dictates Cd4 expression during the development of helper and cytotoxic T cell lineages. *Nature immunology*. 2015 Jul;16(7):746-54.
  61. Viatte S, Plant D, Han B, Fu B, Yarwood A, Thomson W, Symmons DP, Worthington J, Young A, Hyrich KL, Morgan AW. Association of HLA-DRB1 haplotypes with rheumatoid arthritis severity, mortality, and treatment response. *Jama*. 2015 Apr 28;313(16):1645-56.
  62. Viatte S, Plant D, Bowes J, Lunt M, Eyre S, Barton A, Worthington J. Genetic markers of rheumatoid arthritis susceptibility in anti-citrullinated peptide antibody negative patients. *Annals of the rheumatic diseases*. 2012 Dec 1;71(12):1984-90.
  63. Viatte S, Massey J, Bowes J, Duffus K, arcOGEN Consortium, Eyre S, Barton A, Worthington J, Loughlin J, Arden N, Birrell F. Replication of associations of genetic loci outside the hla region with susceptibility to anti-cyclic citrullinated peptide-negative rheumatoid arthritis. *Arthritis & rheumatology*. 2016 Jul;68(7):1603-13.
  64. Han GG, Lee JY, Jin GD, Park J, Choi YH, Kang SK, Chae BJ, Kim EB, Choi YJ. Tracing of the fecal microbiota of commercial pigs at five growth stages from birth to shipment. *Scientific reports*. 2018 Apr 16;8(1):1-9.
  65. Lange CP, Campan M, Hinoue T, Schmitz RF, van der Meulden-de Jong AE, Slingerland H, Kok PJ, van Dijk CM, Weisenberger

- DJ, Shen H, Tollenaar RA. Genome-scale discovery of DNA-methylation biomarkers for blood-based detection of colorectal cancer. *PLoS one*. 2012 Nov 28;7(11):e50266.
66. Nair N, Wilson AG, Barton A. DNA methylation as a marker of response in rheumatoid arthritis. *Pharmacogenomics*. 2017 Sep;18(14):1323-32.
67. Plant D, Webster A, Nair N, Oliver J, Smith SL, Eyre S, Hyrich KL, Wilson AG, Morgan AW, Isaacs JD, Worthington J. Differential methylation as a biomarker of response to etanercept in patients with rheumatoid arthritis. *Arthritis & Rheumatology*. 2016 Jun;68(6):1353-60.
68. Maeshima K, Stanford SM, Hammaker D, Sacchetti C, Zeng LF, Ai R, Zhang V, Boyle DL, Muench GR, Feng GS, Whitaker JW. Abnormal PTPN11 enhancer methylation promotes rheumatoid arthritis fibroblast-like synoviocyte aggressiveness and joint inflammation. *JCI insight*. 2016 May 19;1(7).
69. Svendsen AJ, Gervin K, Lyle R, Christiansen L, Kyvik K, Junker P, Nielsen C, Houen G, Tan Q. Differentially methylated DNA regions in monozygotic twin pairs discordant for rheumatoid arthritis: an epigenome-wide study. *Frontiers in immunology*. 2016 Nov 17;7:510.
70. Karouzakis E, Raza K, Kolling C, Buckley CD, Gay S, Filer A, Ospelt C. Analysis of early changes in DNA methylation in synovial fibroblasts of RA patients before diagnosis. *Scientific reports*. 2018 May 9;8(1):1-6.
71. Svendsen AJ, Gervin K, Lyle R, Christiansen L, Kyvik K, Junker P, Nielsen C, Houen G, Tan Q. Differentially methylated DNA regions in monozygotic twin pairs discordant for rheumatoid arthritis: an epigenome-wide study. *Frontiers in immunology*. 2016 Nov 17;7:510.
72. Tabares P, Berr S, Langenhorst D, Sawitzki B, Ten Berge I, Tony HP, Hünig T. Short-term cytokine stimulation reveals regulatory T cells with down-regulated Foxp3 expression in human peripheral blood. *European journal of immunology*. 2018 Feb;48(2):366-79.
73. Moosavi A, Ardekani AM. Role of epigenetics in biology and human diseases. *Iranian biomedical journal*. 2016 Nov;20(5):246.
74. Available online: [https://www.epigenomics.com/wpcontent/uploads/2016/06/approval\\_pm\\_eng.pdf](https://www.epigenomics.com/wpcontent/uploads/2016/06/approval_pm_eng.pdf) (accessed on 22 August 2016).
75. Nair N, Plant D, Verstappen SM, Isaacs JD, Morgan AW, Hyrich KL, Barton A, Wilson AG, MATURA investigators. Differential DNA methylation correlates with response to methotrexate in rheumatoid arthritis. *Rheumatology*. 2020 Jun 1;59(6):1364-71.