Winter 2020, Volume 5, Issue 16 (3-5)





Effects of iron oxide nanoparticles on MDA-MB-231 breast cancer cell line

Ramadhan Ibrahim¹*, Mohammad Ali Saremi², Mohammad Ali Keshavarz Shahbaz³, Maryam forouhi⁴

¹ Fish Recourses and Aquatic Animals Department, College of Agriculture Salahaddin University, Erbil, Iraq.

²Personalized Medicine Research Center of AmitisGen, Tehran, Iran.

³Human Genetics Division, Medical Biotechnology Department, National Institute of Genetics Engineering and Biotechnology, Tehran, Iran.

⁴National Institute of Genetic Engineering and Biotechnology, Tehran, Iran

*Corresponding author: Ramadhan Ibrahim, Fish Recourses and Aquatic Animals Department, College of Agriculture Salahaddin University, Erbil, Iraq

DOI: 10.22034/pmj.2020.40425

Submitted: 2020/01/13	Abstract
Accepted: 2020/02/19	Because of their small size unique physics, and chemical properties, metal papoparticles
Keywords:	can easily cross obstacles and reach their target cells, which makes them an ideal choice
Iron oxide nanoparticle	for therapeutic purposes in various cancers. In this study, the effects of iron oxide
Oxidative stress	nanoparticles on MDA-MB-231 breast cancer cell line were examined, and biomarkers
reactive oxygen species	related to oxidative stress were evaluated. Fe2O3 nanoparticles were suspended in a cell
Breast cancer	culture medium and diluted to appropriate concentrations (0, 10, 30, 60, and 120 μ g/ml)
	for 24 and 48 h. GSH, superoxide dismutase, catalase, and ROS generation were evaluated.
©2020.Personalized Medicine Journal	The results showed that iron oxide nanoparticles induced intracellular ROS generation in a
	dose- and time-dependent manner. The results further showed that iron oxide nanoparticles
	increased ROS and activated oxidative stress in cells.

INTRODUCTION

Today, the use of metal nanoparticles is one of the most important issues in various medical fields (1). Because of their small size and unique physics and chemical properties, metal nanoparticles can easily cross obstacles and reach their target cells, so they are an ideal choice for therapeutic purposes in various cancers (2). Numerous studies have investigated the side effects of using nanoparticles; however, there are still many unanswered questions about the toxic properties of these substances on biological systems (3). A recent study found that metal nanoparticles increased the production of oxidative stress in cells by increasing the production of reactive oxygen species (ROS) and reducing the activity of the intracellular glutathione system (4). Oxidative stress plays an important role in cellular processes such as cellular signaling, cell proliferation, and genotoxic responses (5). Superoxide dismutase, catalase, and GSH reductase are known natural antioxidants that neutralize excessive ROS and prevent it from damaging the cellular structure (6).

The most common cancers in females worldwide are breast and cervical cancers. An estimated 1 in 8 women in the United States will develop one of these diseases (7). The incidence of breast cancer in developing countries is 23% in young adults (15-49 years) compared with 10% in developed countries (8). In menopausal females (>50 years), the incidence rate is 28% in developing countries compared with 39% in developed countries (9). Iron oxide nanoparticles are of particular interest for in vivo applications, including magnetic resonance imaging for medical diagnosis, hyperthermia in cancer therapy, tissue repair, drug delivery, and cellular therapy (10). In the current study, the effects of iron oxide nanoparticles on a MDA-MB-231 breast cancer cell line were examined, and the biomarkers related to oxidative stress, including GSH, superoxide dismutase, catalase, and ROS generation, were evaluated.

MATERIALS AND METHODS

Human breast cancer cells (MDA-MB-231) were procured from the Pasteur Institute of Iran. They were preserved and subcultured in the laboratory and used to determine cytotoxicity against iron oxide nanoparticles. Iron oxide (Fe2O3) nanoparticles were suspended in the cell culture medium and diluted to appropriate concentrations (0, 10, 30, 60, and 120 μ g/ml) for 24 and 48 h. Cells not exposed to iron oxide nanoparticles served as the control in each experiment. GSH levels were quantified by using Ellman's reagent (DTNB). The reaction was monitored at 412 nm, and the amount of GSH was expressed in terms of nanomoles of GSH per milligram of protein. Superoxide Dismutase was assayed by NTB buffer, and the absorbance of the reaction mixture was measured at 450 nm. Catalase

Copyright © The Author(s). This is an open-access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, p rovided the original work is properly cited.

activity was measured by its ability to split hydrogen peroxide (H2O2) within 1 min of incubation time. Catalase activity was expressed as micromoles of H2O2 decomposed per minute per milligram of protein. ROS generation was assessed in MDA-MB-231 cells using DCFH-DA dye as the fluorescence agent. Data was expressed as mean (\pm SE) and analyzed by one-way analysis of variance (ANOVA). A *p*-value <0.05 was considered statistically significant.

RESULTS

The ability of iron oxide nanoparticles to induce oxidative stress was assayed by measuring levels of GSH, ROS, superoxide dismutase, and catalase in a MDA-MB-231 breast cancer cell line. The results showed that iron oxide nanoparticles induced intracellular ROS generation in a dose- and timedependent manner (Table 1).

Table1. Effect of treatment of different doses of iron oxide nanoparticles at different times on oxidative stress biomarkers of MDA-MB-231 breast cancer cell line

Stress oxidative biomarker	ROS (DCF-fluorescence)		GSH (nMcSH/mg)		SOD (Unit/ml)		Catalase (% of control)	
Treatment time	24h	48h	24h	48h	24h	48h	24h	48h
Control	100	102	14.5	14.6	2.1	2.2	98	99
10 conc	110	115	14	13.8	1.9	1.9	100	1.5
30 conc	125	150	13.5	13	1.8	1.5	97	88
60 conc	170	180	12	11	1.5	1.3	87	75
120 conc	200	230	9	5	1.4	1.0	68	60

DISCUSSION

Iron oxide nanoparticles are widely used in a variety of medical fields, from magnetic resonance imaging to drug design. Still, there are concerns about the toxic properties and effects their use can have on the environment (11). This study evaluated the effect of this substance on the oxidative stress of the MDA-MB-231 breast cancer cell line. The results showed that iron oxide nanoparticles can increase the oxidative stress activity of breast cancer cells by inducing the production of free radicals; thus, they are cytotoxic. Previous studies have shown that nanoparticles of various sizes and different chemical compositions attack mitochondria, which are redoxactive organelles. Mitochondria are a prominent site of ROS formation in cells exposed to nanoparticles. Therefore, nanoparticles may change the production of ROS and affect antioxidant defenses to induce oxidative stress (12). By increasing the dose of iron oxide nanoparticles and increasing the treatment time, this study showed that ROS was increased and the values of GSH, SOD, and catalase were decreased significantly, indicating an increase in cell stress and a decrease in the cell antioxidant system. ROS typically include the superoxide radical (O2–), hydrogen peroxide (H2O2), and the hydroxyl radical (OH), which cause damage to cellular components including DNA damage and ultimately apoptotic cell death (13). Previous studies have shown that ROS are involved in damaging DNA, causing damage to both purine and pyrimidine bases as well as the DNA backbone. The current results showed that iron oxide nanoparticles increased ROS and activate oxidative stress in cells.

REFERENCE

1. Alghamdi IG, Hussain II, AlghamdiMS, El-SheemyMA (2013) The incidence rate of female breast cancer in Saudi Arabia: an observational descriptive epidemiological analysis of data from Saudi Cancer Registry 2001-2008. Breast Cancer Targets Ther 5:103–109 2. Jain TK, Reddy MK,Morales MA, Leslie-Pelecky DL, Labhasetwar V (2008) Biodistribution, clearance, and biocompatibility of iron oxide magnetic nanoparticles in rats. Mol Pharm 5(2):316–327

3. Lee KJ, An JH, Shin JS, Kim DH (2012) Synthesis and characterization of bicalutamide-loaded magnetic nanoparticles as anti-tumor drug carriers. J Nanosci Nanotechnol 12(2):1611–1615 4. Maeng JH, Lee DH, Jung KH, Bae YH, Park IS, Jeong S, Jeon YS, Shim CK, KimW, Kim J, Lee J, Lee YM, Kim JH, KimWH, Hon SS (2010) Multi-functional doxorubicin-loaded super paramagnetic iron oxide nanoparticles for chemotherapy and magnetic resonance imaging in liver cancer. Biomaterials 31(18):4995–5006

5. Jia Y, YuanM, Yuan H,Huang X, Sui X, Cui X, Tang F, Peng J, Chen J, Lu S,XuW, Zhang L,GuoQ (2012) Co-encapsulation ofmagnetic Fe3O4 nanoparticles and doxorubicin into biodegradable PLGA nanocarriers for intratumoral drug delivery. Int J Nanomedicine 63: 1697–1708

6. Li Y-F, Chen C (2011) Fate and toxicity of metallic and metalcontaining nanoparticles for biomedical applications. Small 7: 2965–2980

7. Singh N, Manshian B, Jenkins GJS, Griffiths SM, Williams PM, Maffeis TG, Wright CJ, Doak SH (2009) Nano genotoxicology: the DNA damaging potential of engineered nanomaterials. Biomaterials 30:3891–3914

8. Skocaj M, FilipicM, Petkovic J, Novak S (2011) Titaniumdioxide in our everyday life; is it safe? Radiol Oncol 45:227–247

9. Wang Y, AkerWG, Hwang HM, Yedjou CGYH et al (2011) A study of themechanism of in vitro cytotoxicity ofmetal oxide nanoparticles using catfish primary hepatocytes and human HepG2 cells. Sci Total Environ 409:4753–4762

10. van Maanen JM, Borm PJ, Knaapen A, van Herwijnen M, Schilderman PA, Smith KR, Aust AE, Tomatis M, Fubini B (1999)

In vitro effects of coal fly ashes: hydroxyl radical generation, iron release, and DNA damage and toxicity in rat lung epithelial cells. Inhal Toxicol 11:1123–1141

 Borm PJ, Schins RP, Albrecht C (2004) Inhaled particles and lung cancer, part B: paradigms and risk assessment. Int J Cancer 110:3–14
Naziroglu M, Simsek M, Kutlu M (2004) Moderate exercise with dietary vitamin C and E combination protects streptozotocin induced oxidative damage to the blood and improves fetal outcomes in pregnant rats. Clin Chem Lab Med 42:511–517

13. Mossman T (1983) Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. J Immunol Methods 65:55–63